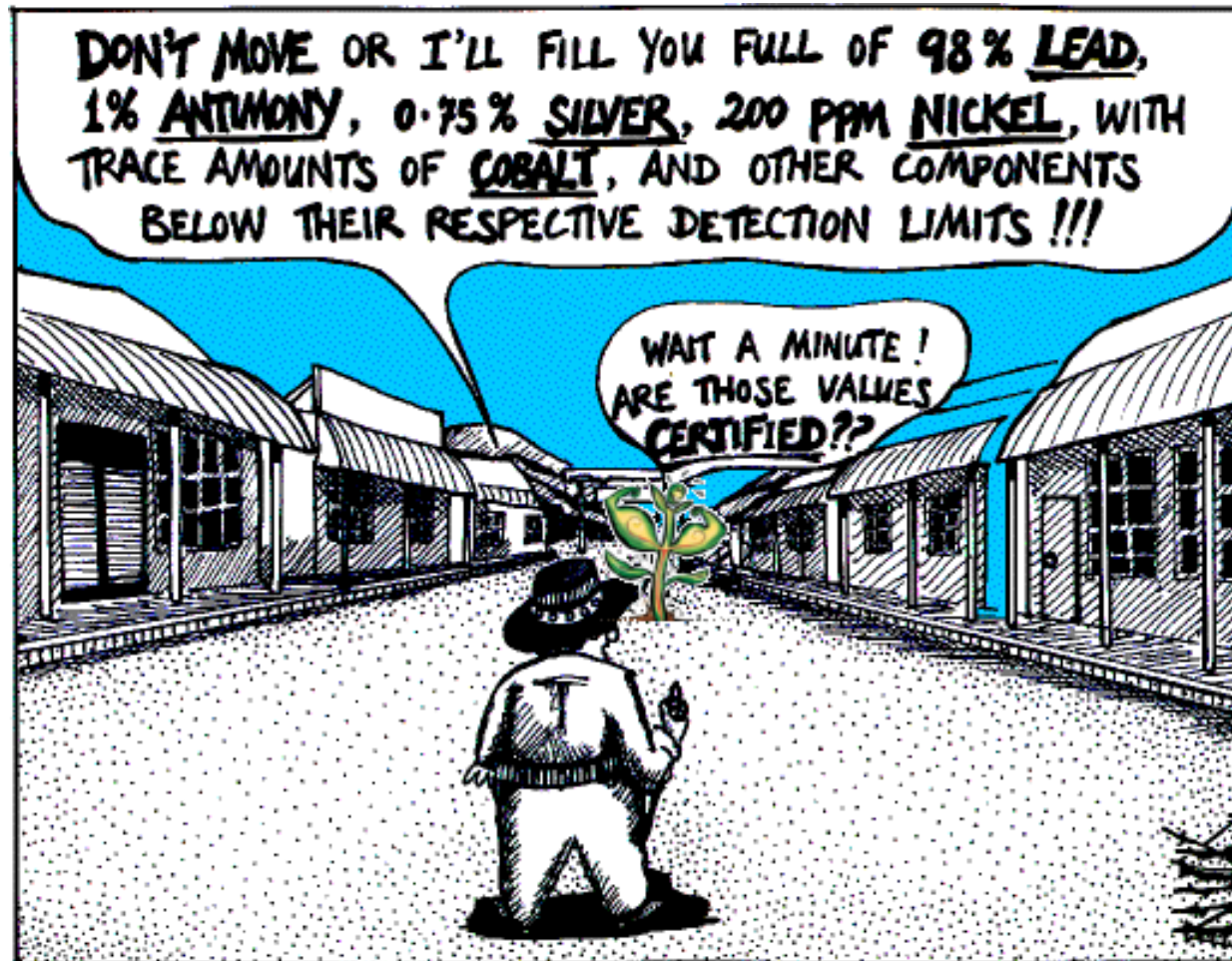


# Heavy Metals and Plants - a complicated relationship

## → Cadmium toxicity



Heavy metal-hyperaccumulation in the Wild West

modified from: <http://strangematter.sci.waikato.ac.nz/>

Hendrik Küpper based on a talk of Elisa Andresen, Advanced Course on Bioinorganic Chemistry & Biophysics of Plants, summer semester 2019

# Cadmium

1	1 <b>H</b> 1.008																	2 <b>He</b> 4.0026		
2	3 <b>Li</b> 6.94	4 <b>Be</b> 9.0122													5 <b>B</b> 10.81	6 <b>C</b> 12.011	7 <b>N</b> 14.007	8 <b>O</b> 15.999	9 <b>F</b> 18.998	10 <b>Ne</b> 20.180
3	11 <b>Na</b> 22.990	12 <b>Mg</b> 24.305													13 <b>Al</b> 26.982	14 <b>Si</b> 28.085	15 <b>P</b> 30.974	16 <b>S</b> 32.06	17 <b>Cl</b> 35.45	18 <b>Ar</b> 39.948
4	19 <b>K</b> 39.098	20 <b>Ca</b> 40.078	21 <b>Sc</b> 44.956	22 <b>Ti</b> 47.867	23 <b>V</b> 50.942	24 <b>Cr</b> 51.996	25 <b>Mn</b> 54.938	26 <b>Fe</b> 55.845	27 <b>Co</b> 58.933	28 <b>Ni</b> 58.693	29 <b>Cu</b> 63.546	30 <b>Zn</b> 65.38	31 <b>Ga</b> 69.723	32 <b>Ge</b> 72.63	33 <b>As</b> 74.922	34 <b>Se</b> 78.96	35 <b>Br</b> 79.904	36 <b>Kr</b> 83.798		
5	37 <b>Rb</b> 85.468	38 <b>Sr</b> 87.62	39 <b>Y</b> 88.906	40 <b>Zr</b> 91.224	41 <b>Nb</b> 92.906	42 <b>Mo</b> 95.96	43 <b>Tc</b> [97.91]	44 <b>Ru</b> 101.07	45 <b>Rh</b> 102.91	46 <b>Pd</b> 106.42	47 <b>Ag</b> 107.87	48 <b>Cd</b> 112.41	49 <b>In</b> 114.82	50 <b>Sn</b> 118.71	51 <b>Sb</b> 121.76	52 <b>Te</b> 127.60	53 <b>I</b> 126.90	54 <b>Xe</b> 131.29		
6	55 <b>Cs</b> 132.91	56 <b>Ba</b> 137.33	* 71 <b>Lu</b> 174.97	72 <b>Hf</b> 178.49	73 <b>Ta</b> 180.95	74 <b>W</b> 183.84	75 <b>Re</b> 186.21	76 <b>Os</b> 190.23	77 <b>Ir</b> 192.22	78 <b>Pt</b> 195.08	79 <b>Au</b> 196.97	80 <b>Hg</b> 200.59	81 <b>Tl</b> 204.38	82 <b>Pb</b> 207.2	83 <b>Bi</b> 208.98	84 <b>Po</b> [208.98]	85 <b>At</b> [209.99]	86 <b>Rn</b> [222.02]		
7	87 <b>Fr</b> [223.02]	88 <b>Ra</b> [226.03]	** 103 <b>Lr</b> [262.11]	104 <b>Rf</b> [265.12]	105 <b>Db</b> [268.13]	106 <b>Sg</b> [271.13]	107 <b>Bh</b> [270]	108 <b>Hs</b> [277.15]	109 <b>Mt</b> [276.15]	110 <b>Ds</b> [281.16]	111 <b>Rg</b> [280.16]	112 <b>Cn</b> [285.17]	113 <b>Uut</b> [284.18]	114 <b>Fl</b> [289.19]	115 <b>Uup</b> [288.19]	116 <b>Lv</b> [293]	117 <b>Uus</b> [294]	118 <b>Uuo</b> [294]		
*Lanthanoids			* 57 <b>La</b> 138.91	58 <b>Ce</b> 140.12	59 <b>Pr</b> 140.91	60 <b>Nd</b> 144.24	61 <b>Pm</b> [144.91]	62 <b>Sm</b> 150.36	63 <b>Eu</b> 151.96	64 <b>Gd</b> 157.25	65 <b>Tb</b> 158.93	66 <b>Dy</b> 162.50	67 <b>Ho</b> 164.93	68 <b>Er</b> 167.26	69 <b>Tm</b> 168.93	70 <b>Yb</b> 173.05				
**Actinoids			** 89 <b>Ac</b> [227.03]	90 <b>Th</b> 232.04	91 <b>Pa</b> 231.04	92 <b>U</b> 238.03	93 <b>Np</b> [237.05]	94 <b>Pu</b> [244.06]	95 <b>Am</b> [243.06]	96 <b>Cm</b> [247.07]	97 <b>Bk</b> [247.07]	98 <b>Cf</b> [251.08]	99 <b>Es</b> [252.08]	100 <b>Fm</b> [257.10]	101 <b>Md</b> [258.10]	102 <b>No</b> [259.10]				

# Cadmium in the environment

- Rather rare element in Earth's crust (0.1 - 0.5 ppm)
- Some natural sites, associated with ZnS
- Anthropogenic contamination, e.g. ore mining, some fertilizers, car traffic, cigarette smoke, industrial waste, NiCd-batteries

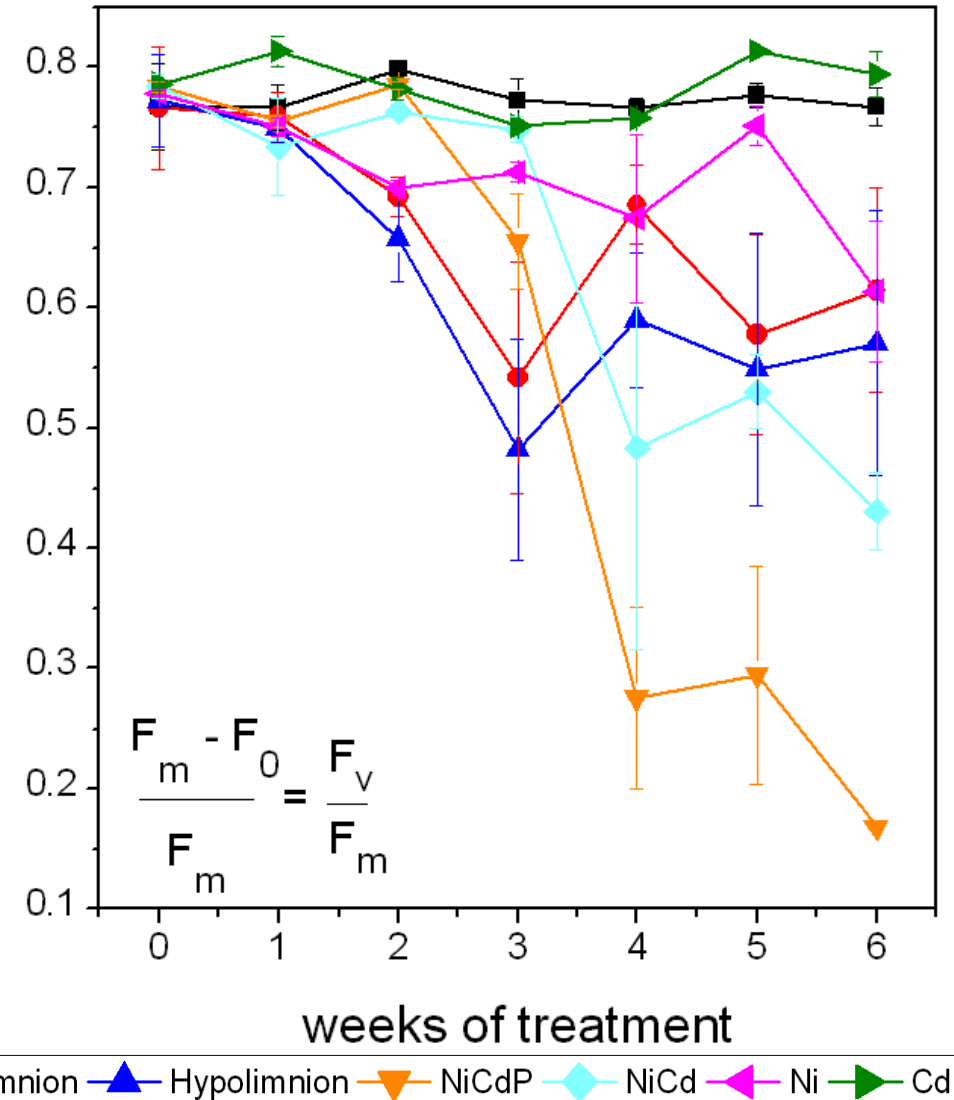
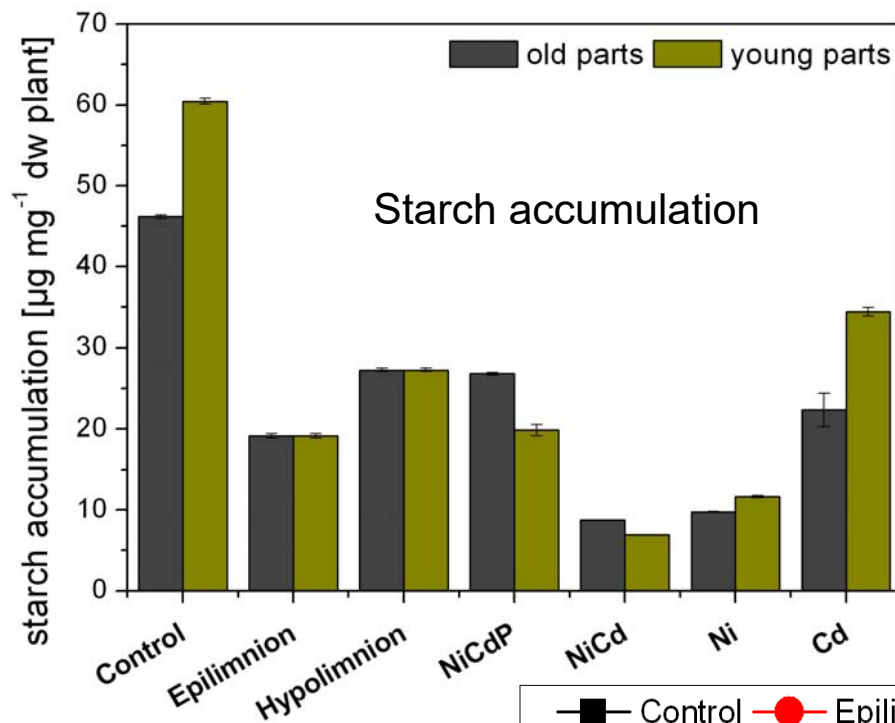


## Cd toxicity – prominent diseases

- **Itai-itai disease** (japanese ouch-ouch sickness)
- 1 of the 4 big pollution diseases in Japan
- Mass cadmium poisoning in Japan, cadmium release into rivers by mining
- Severe pains in joints and spine, softening of the bones, kidney failure
- The mining companies were successfully sued for the damage

# Heavy metal toxicity induced inhibition of photosynthesis at nanomolar concentrations

- Ceratophyllum demersum* plants treated with natural or simulated lake water containing **only 3 nM Cd<sup>2+</sup> and 300 nM Ni<sup>2+</sup>** already show inhibition
- inhibition by Ni+Cd combination treatment much stronger than by the single metals → **synergistic effect!**



# Growth inhibition

Before treatment start



After 1 week



After 2 weeks



After 3 weeks



After 4 weeks

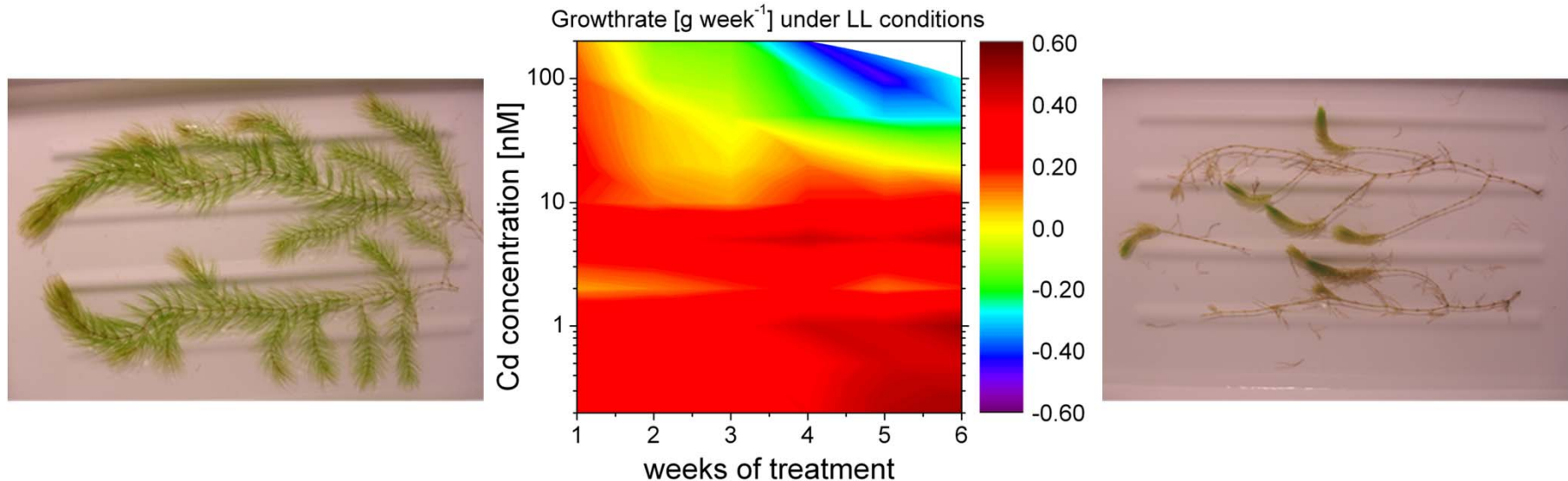
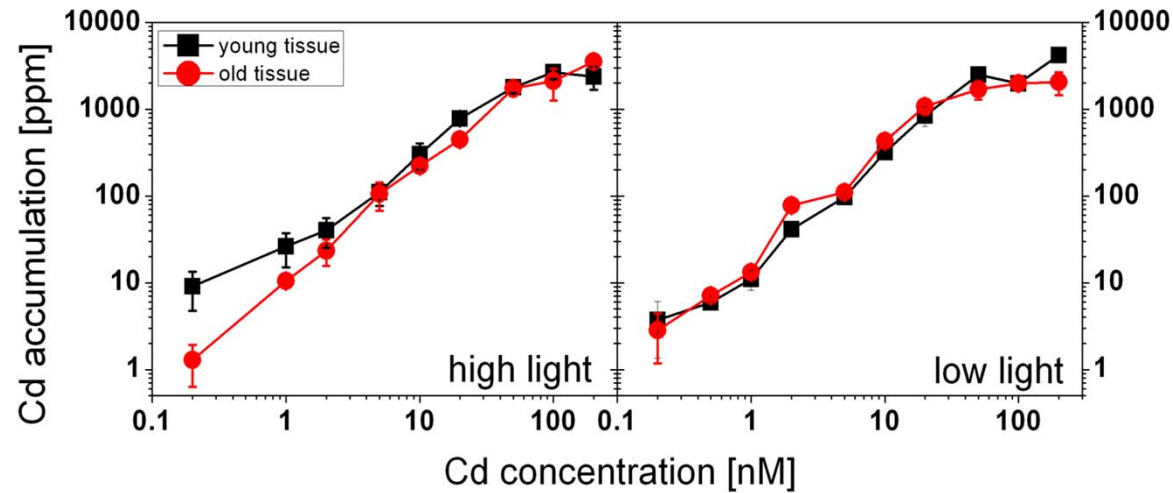


*C. demersum* treated with 200 nM Cd for 4 weeks

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.



# Accumulation of Cd and growth



→ Already low nanomolar Cd is toxic for sensitive plants

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.

# Cd in plants

## Toxicity

- 1) Roots
- 2) Photosynthesis
- 3) Reactive oxygen species
- 4) Genotoxicity
- 5) Stress prevention

Beneficial effects ?!



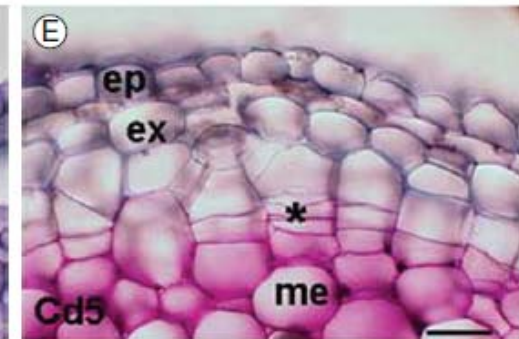
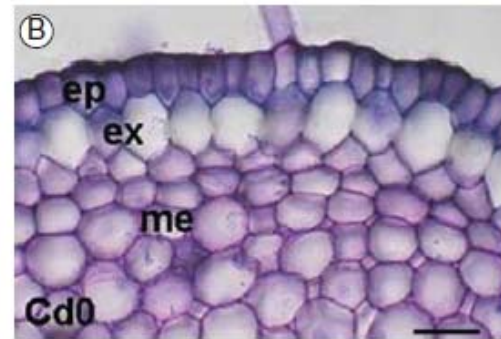
# Cadmium toxicity in plants – 1: Roots

First organ which gets affected

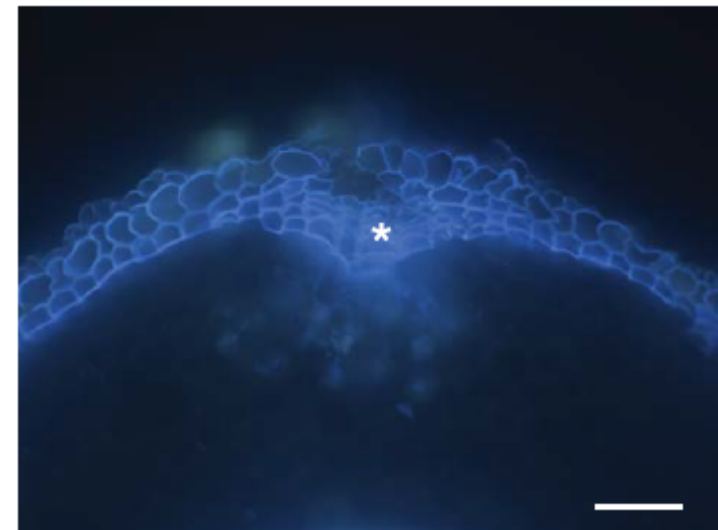
Reduced growth after Cd treatment



More layers of hypodermal periderm



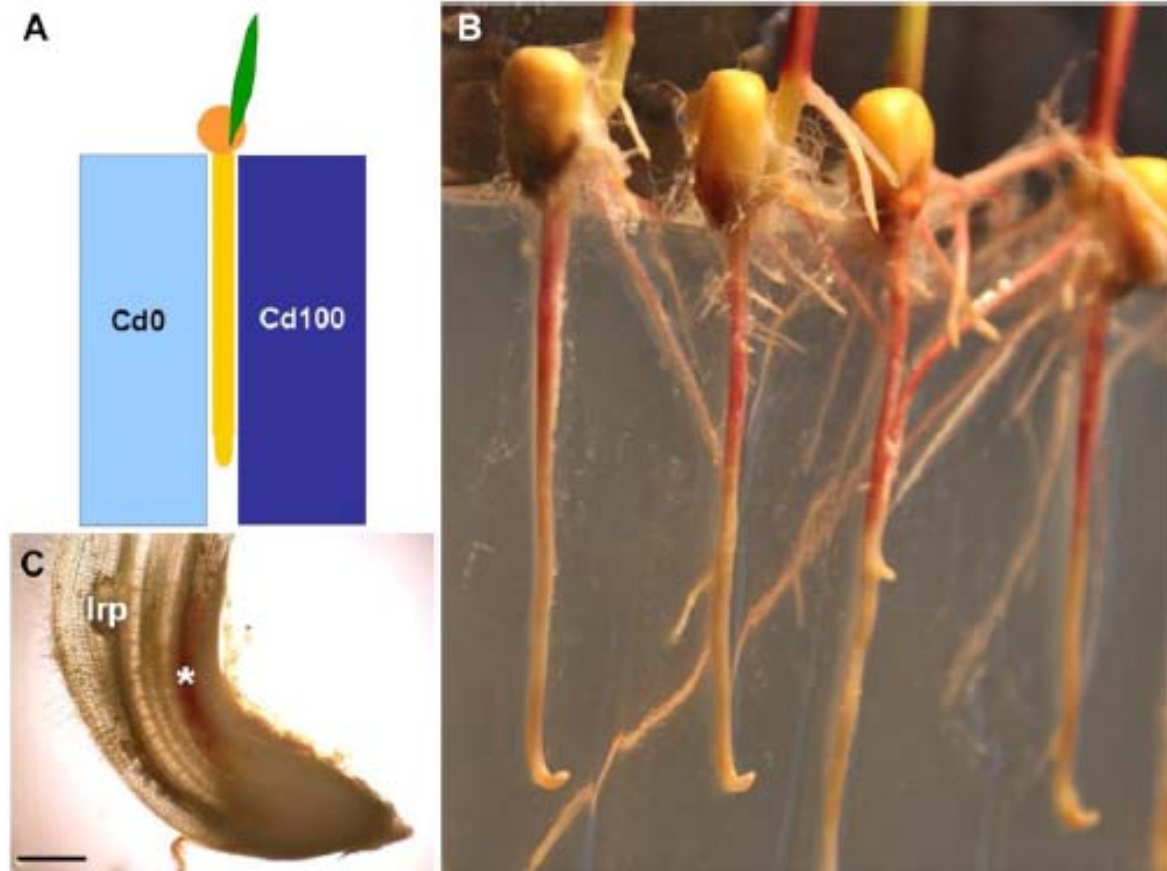
More layers & suberized cell walls (\*) like after injury of root surface



- Maize seedlings with proper roots placed between 2 agar blocks
- one of which contained Cd (50 or 100  $\mu\text{M}$ ), grown in phytochamber under nature-like conditions

→ Roots bending towards the Cd-containing agar → due to growth stop on the Cd-side & continued growth on control-side

Lignification on Cd-exposed side  
(\*) and initiation of lateral root primordium (lrp)

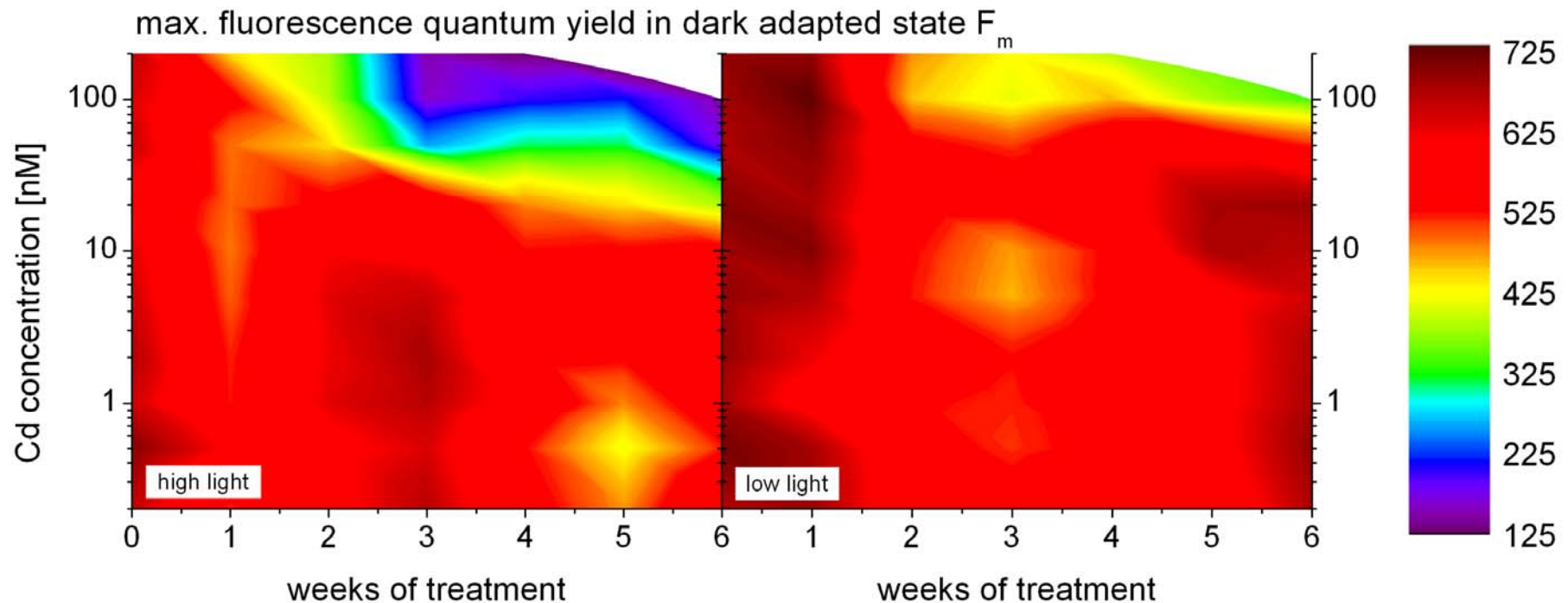


# Cadmium toxicity in plants –

## 2: Photosynthesis

- Indirect measurement: Growth, O<sub>2</sub> production / CO<sub>2</sub> consumption
  - Diminishing the Chl/pigment/protein content
- Direct: Photosynthetic parameters via Chlorophyll fluorescence measurement

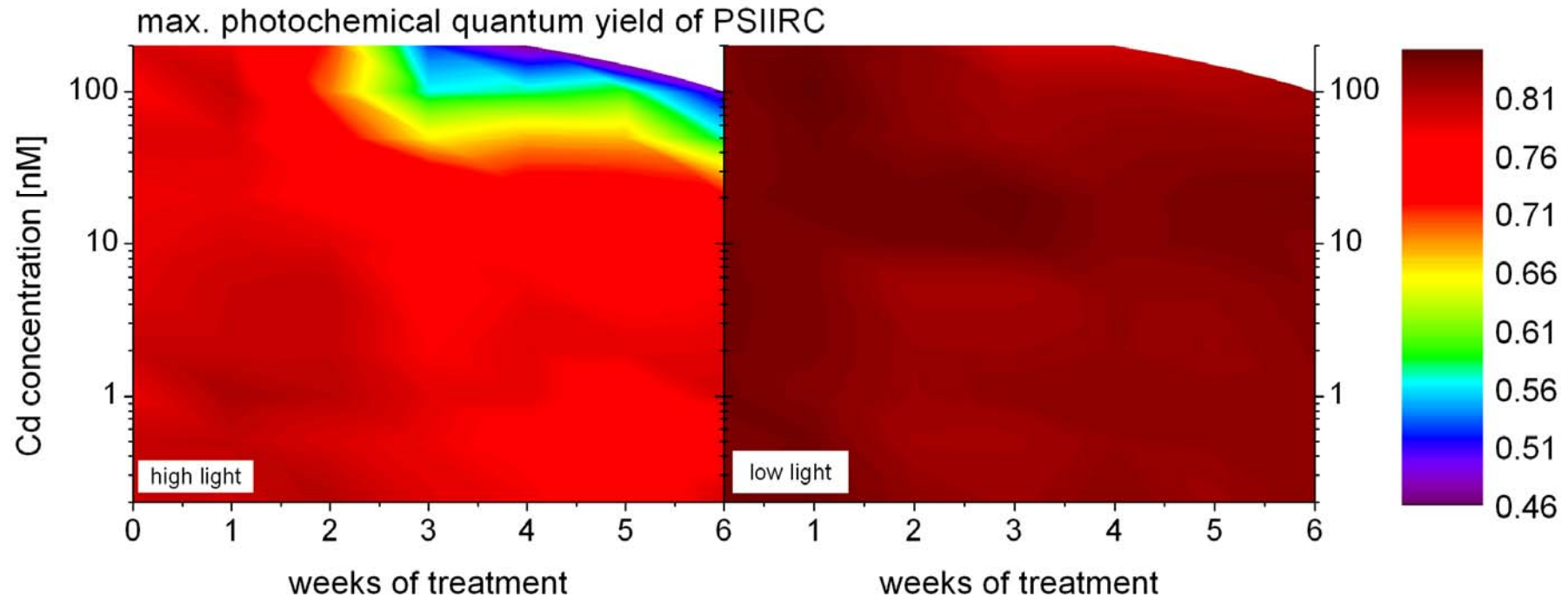
# Cd affects Photosynthesis in *C. demersum*



- Max. fluorescent quantum yield of PSII → amount of fluorescent molecules → LL plants have bigger antenna systems
- Reduced  $F_m$  towards higher concentrations and longer treatment duration → decreased Chl content
- Reduction in week 1 due to acclimation to Cd + HL

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.

# Cd affects Photosynthesis in *C. demersum*



Reduced variable fluorescence  $F_v/F_m = (F_m - F_0) / F_m \rightarrow$  decreased activity of PS II towards higher concentrations and longer treatment duration

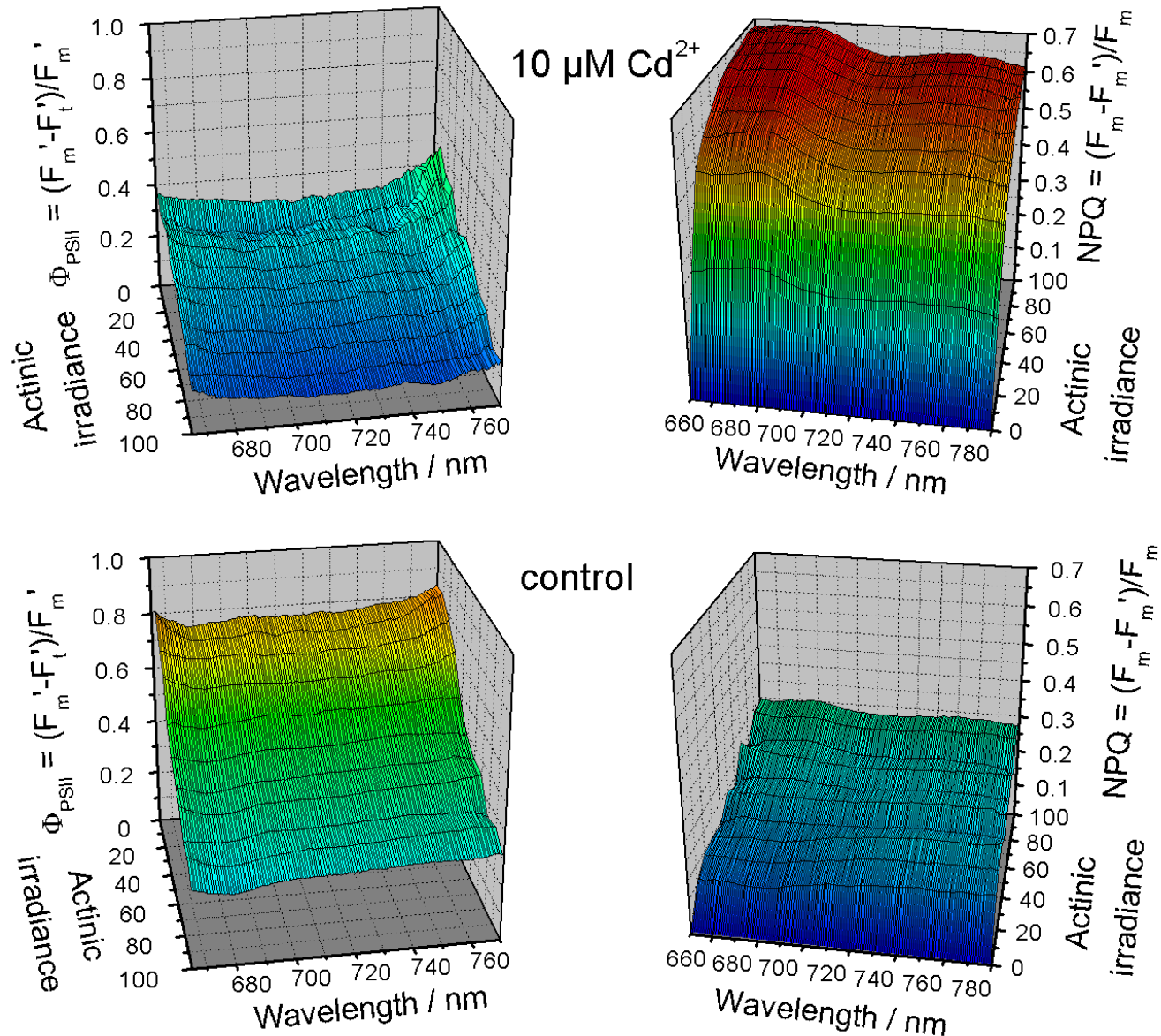
HL much more affected than LL

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.



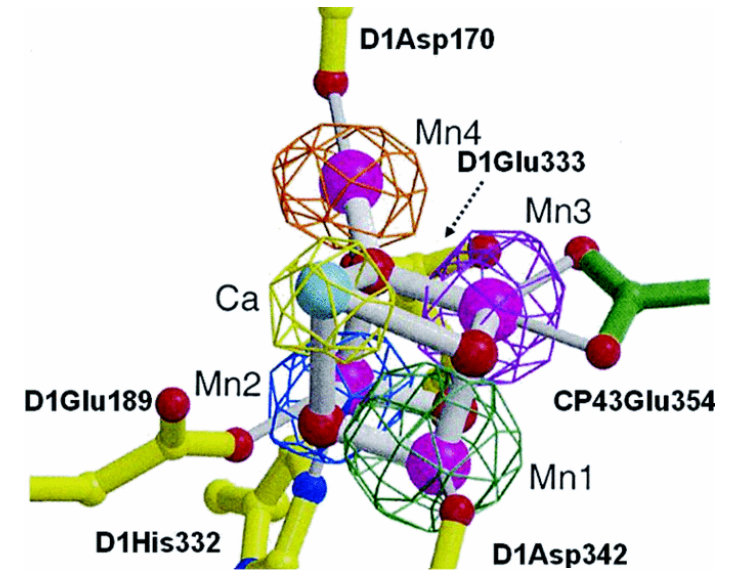
# Cd-stress in the Zn-/Cd-hyperaccumulator *T. caerulescens*:

## Spectral changes of PSII activity parameters

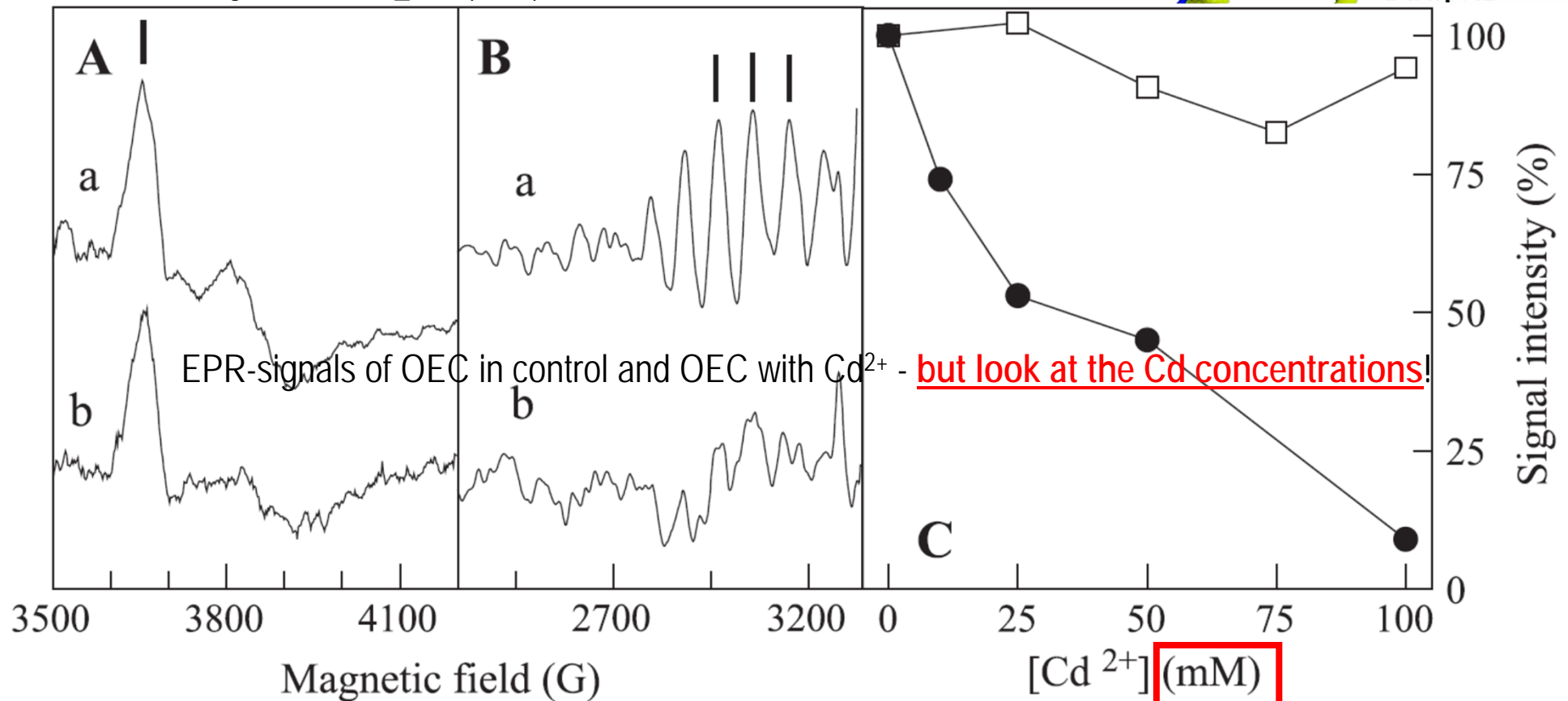




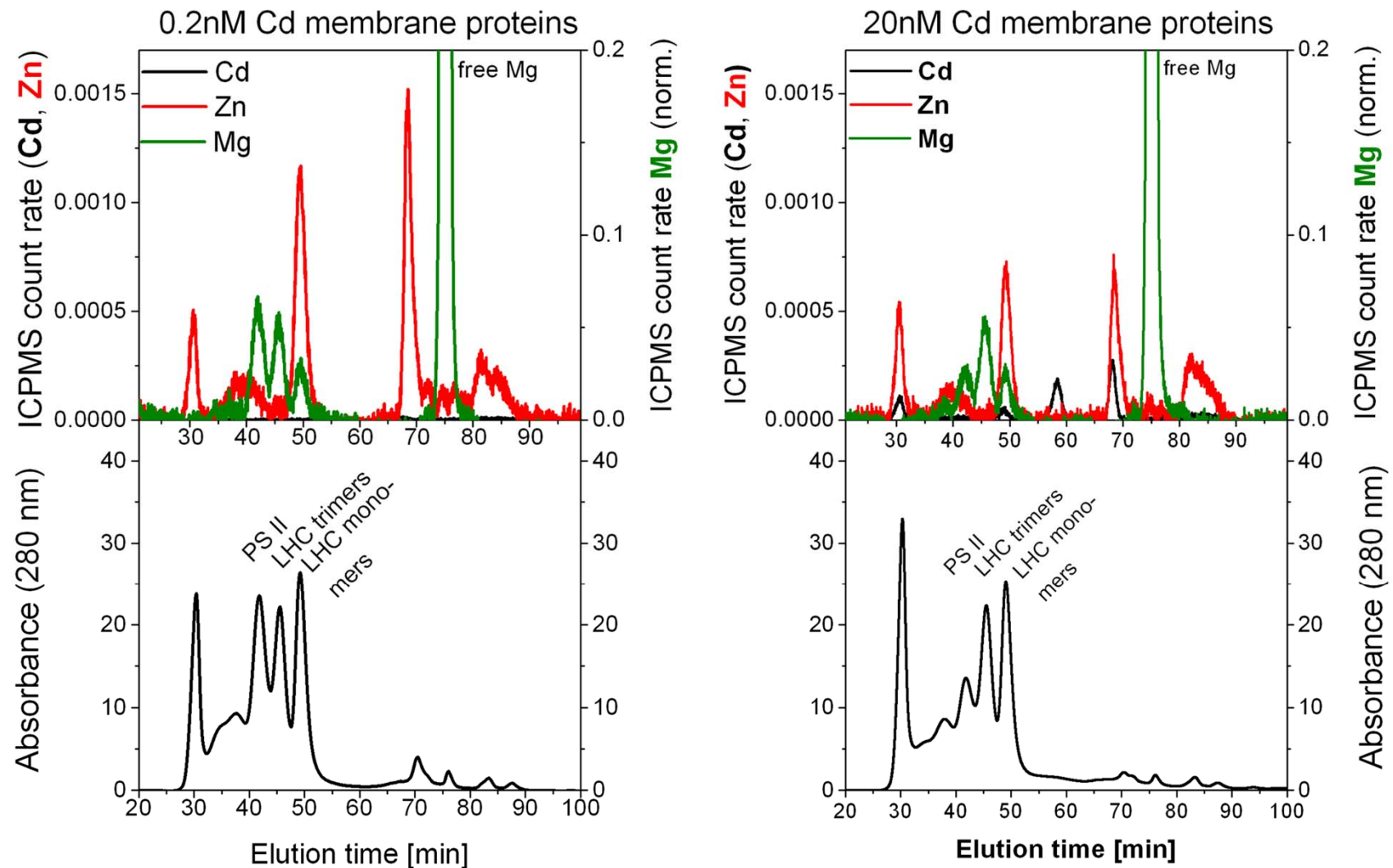
**Substitution of Ca by Cd in water  
splitting complex of PSII  
would inhibit water splitting, if it would  
occur at relevant Cd concentrations**



Sigfridsson KGV\_et al (2004) BBA-Bioen1659, 19-31



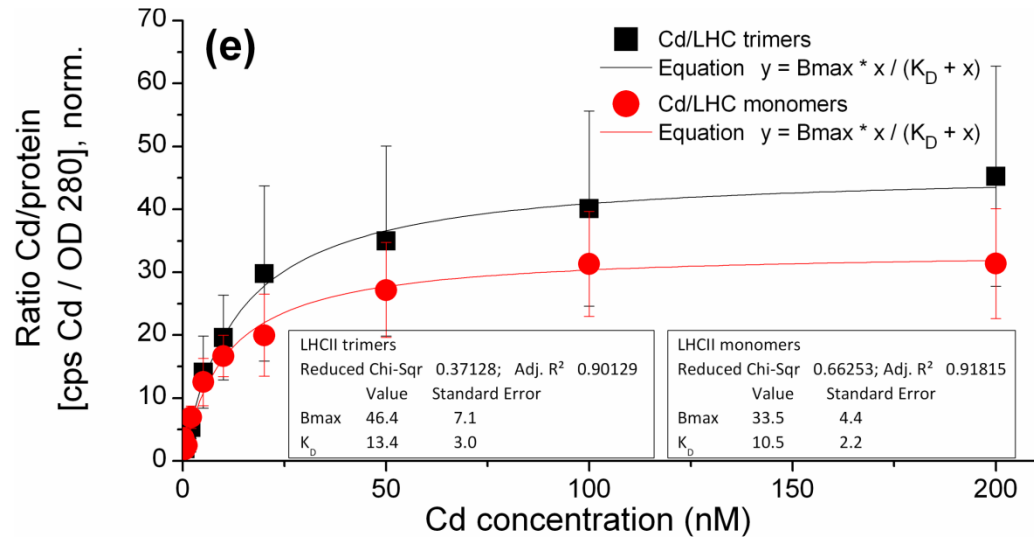
# Cd stress in *C. demersum*: Incorporation of Cd into proteins in LL analysed by metalloproteomics via HPLC-ICP-MS



→ Cd binding to main light harvesting antenna LHCII in LL

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.

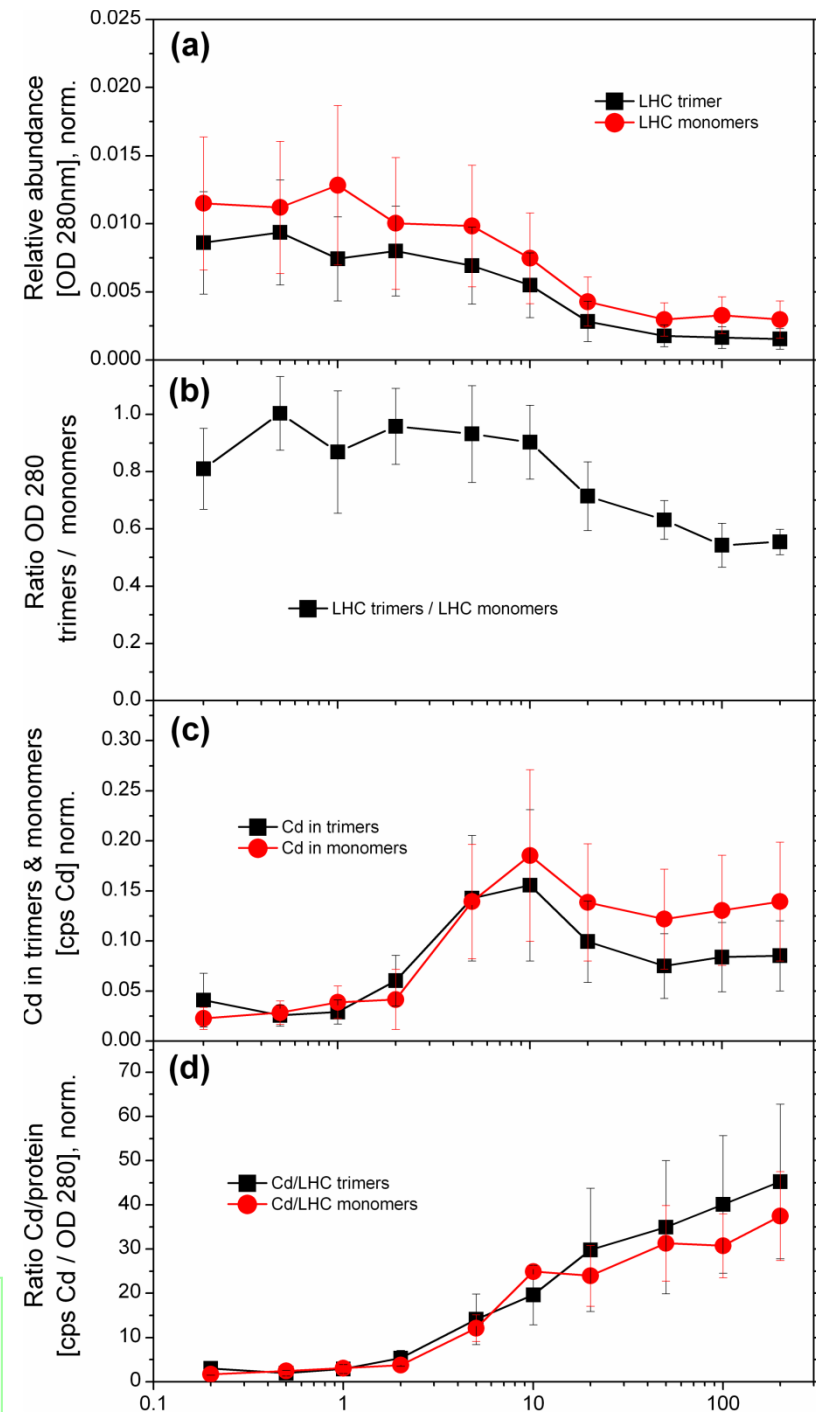
# Example of metal toxicity in the nanomolar range in „normal“ plants: Incorporation of Cd into LHCII in LL



→ Cd binding to LHCII causes disintegration of trimers

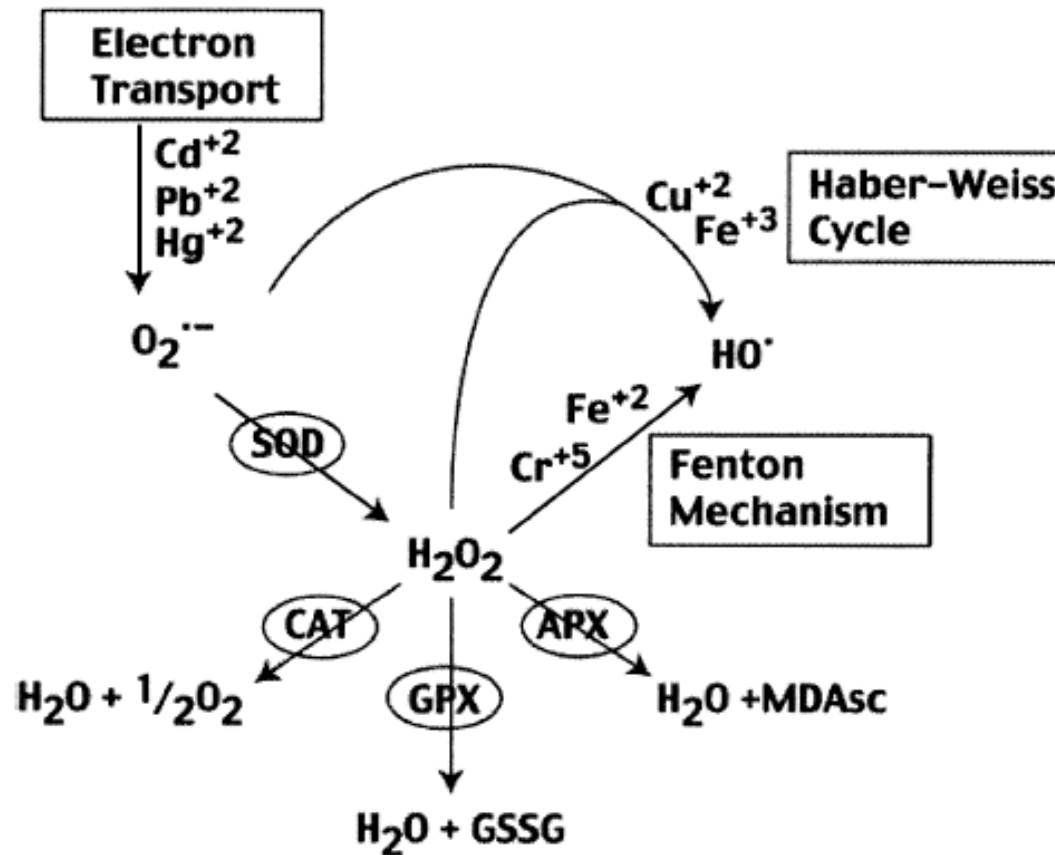
→ Cd bind to LHCII with dissociation constants in the low nanomolar range  
→ diminished photosynthesis despite functional reaction centres!

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.

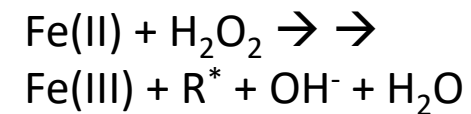


# ROS and Cadmium

- Cadmium redox inert → No Fenton reaction!



Fenton:

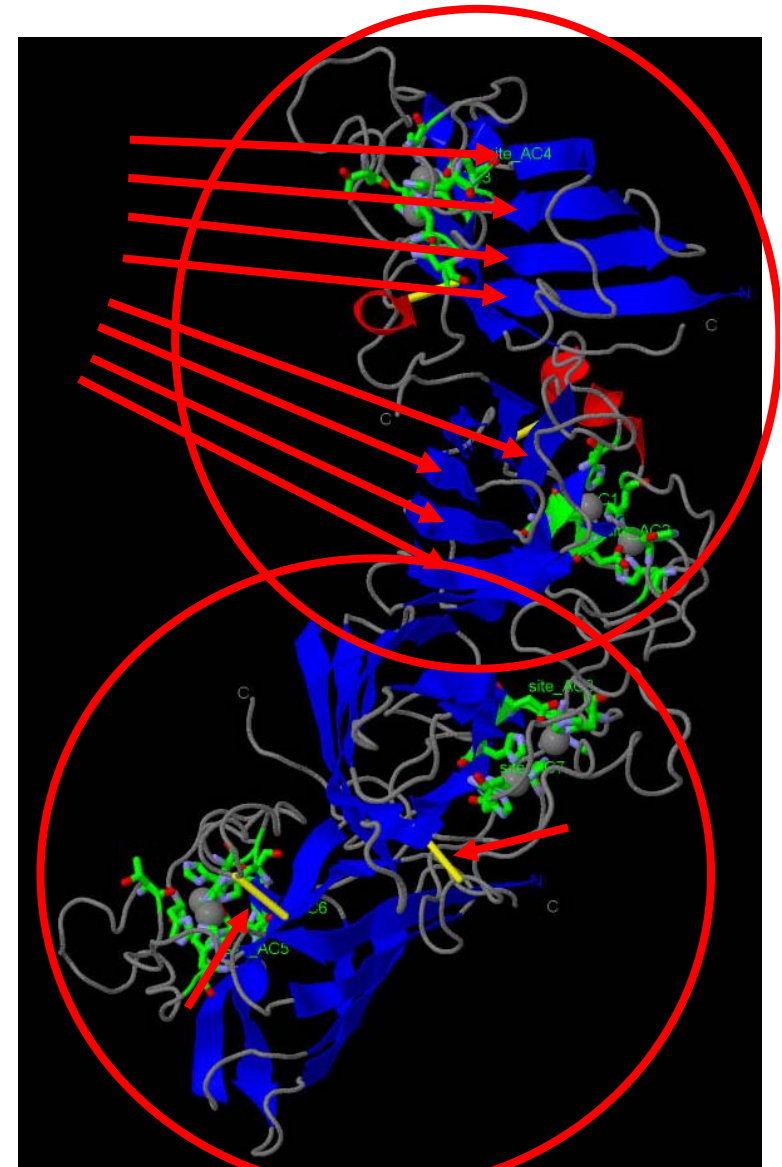
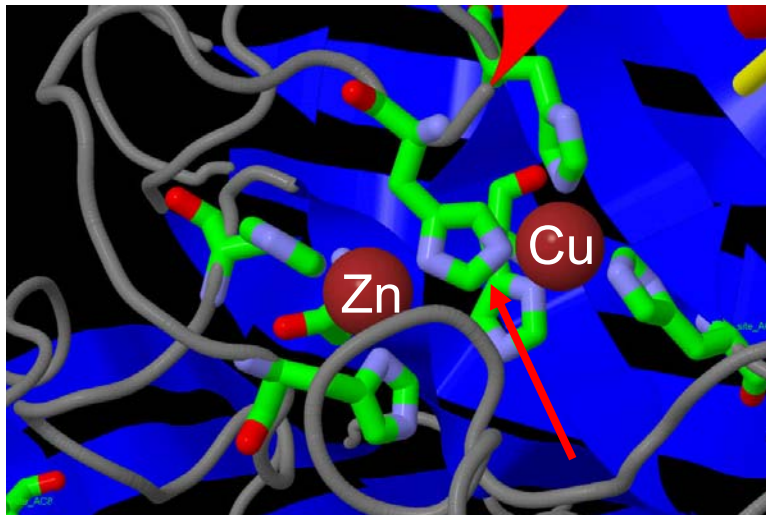


„Biggest source of ROS in animal cells“ ...

→ NEVER shown *in vivo*!

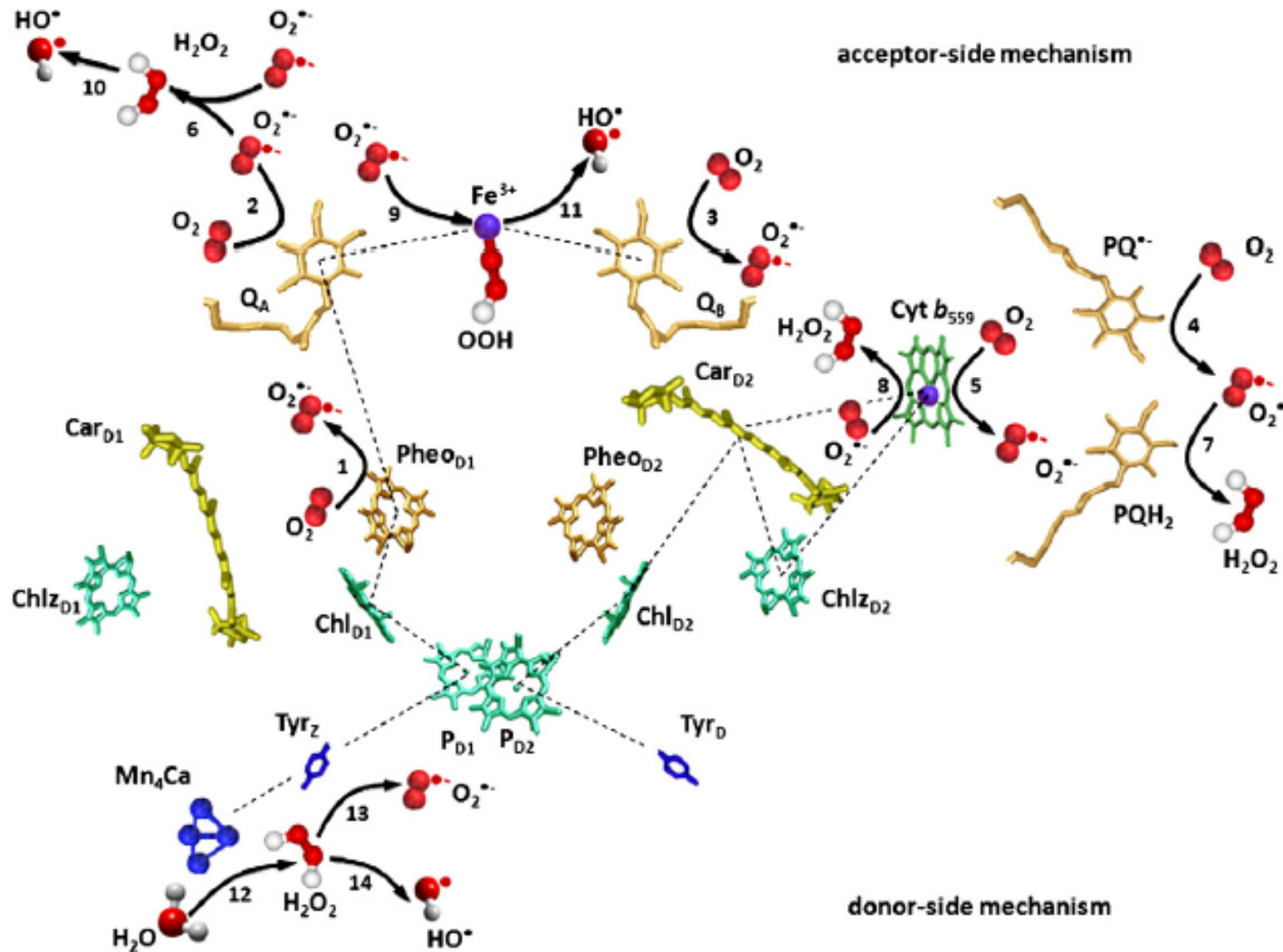
# Possible Target for Cd toxicity: Superoxide dismutase (SOD), in plants a Cu/Zn enzyme

Substitution of Zn by Cd in SOD may contribute to oxidative stress during Cd toxicity



Spinach SOD, From: Kitagawa Y et al., 1991, J Biochem 109, 477-85, images generated with Jena 3D viewer

# Photosynthesis-related ROS





# ROS and Cadmium

ROS production ↑

- Cd interferes with photosynthesis / respiration  
→ electrons transferred to  $O_2$

- In response antioxidant enzymes ↑

Removal of ROS ↓

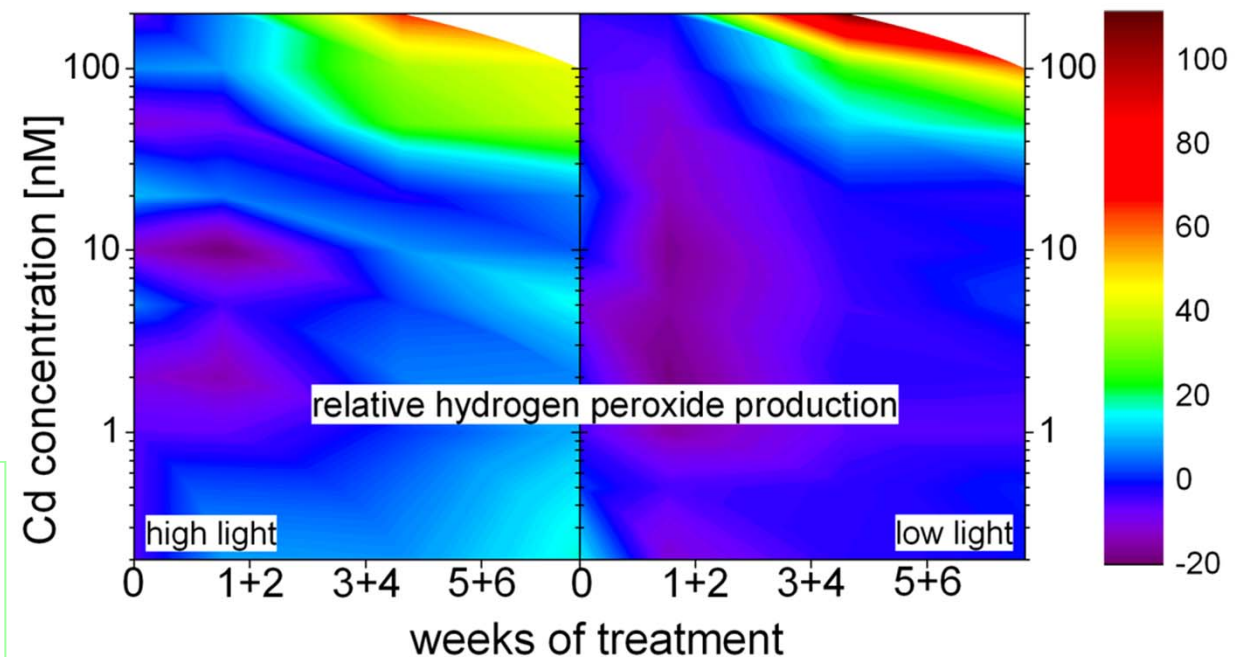
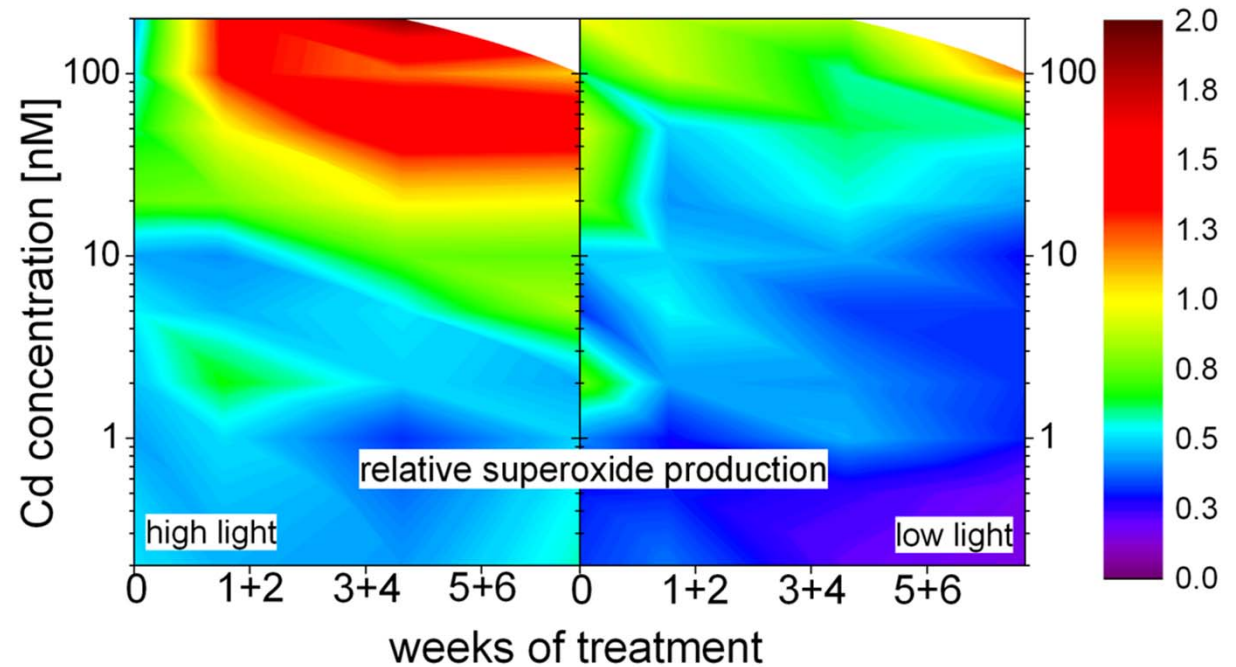
- Cd replaces Zn in SOD (e.g.)  
→ less functional SOD

# Production of reactive oxygen species (ROS) during Cd-stress in HL vs. LL

→ Cd-induced formation of ROS stronger in HL

→ superoxide formation starts at lower Cd concentrations than peroxide formation

Andresen E, Kappel S, Stärk HJ, Riegger U, Borovec J, Mattusch J, Heinz A, Schmelzer CEH, Matoušková Š, Dickinson B, Küpper H (2016) New Phytologist 210, 1244-1258.



## Influence on antioxidant enzymes

- **Lower** Cd concentrations and **shorter** treatment duration tend to increase the antioxidant system
- **Longer** exposure and **higher** Cd concentrations lead to decreased activity or content of the antioxidants

# Cadmium toxicity in plants –

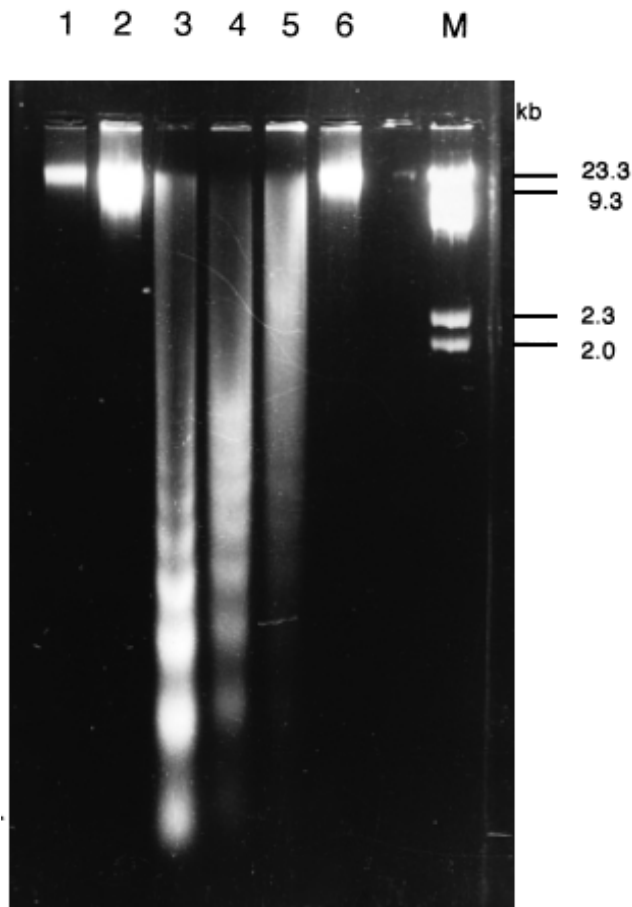
## 4. Genotoxicity

- Induction of DNA damage by
  - direct interaction with the nucleotides
    - modifications like base and sugar lesions, DNA strand breaks, destruction of DNA-protein crosslinks etc.
  - inhibiting DNA repairing enzymes
  - Induction of ROS, ROS lead to lipid peroxidation, which causes membrane damage and production of mutagenic aldehydes

# Methods to detect Genotoxicity

- DNA Analyses
  - Gelelectrophoresis and Comet Assay
  - Random amplification of polymorphism DNA (RAPD)
- DNA / Chromosome Analyses
  - Micronuclei formation
  - Sister chromatid exchange
  - Chromosomal aberrations
- Upregulation of DNA-related / repairing enzymes

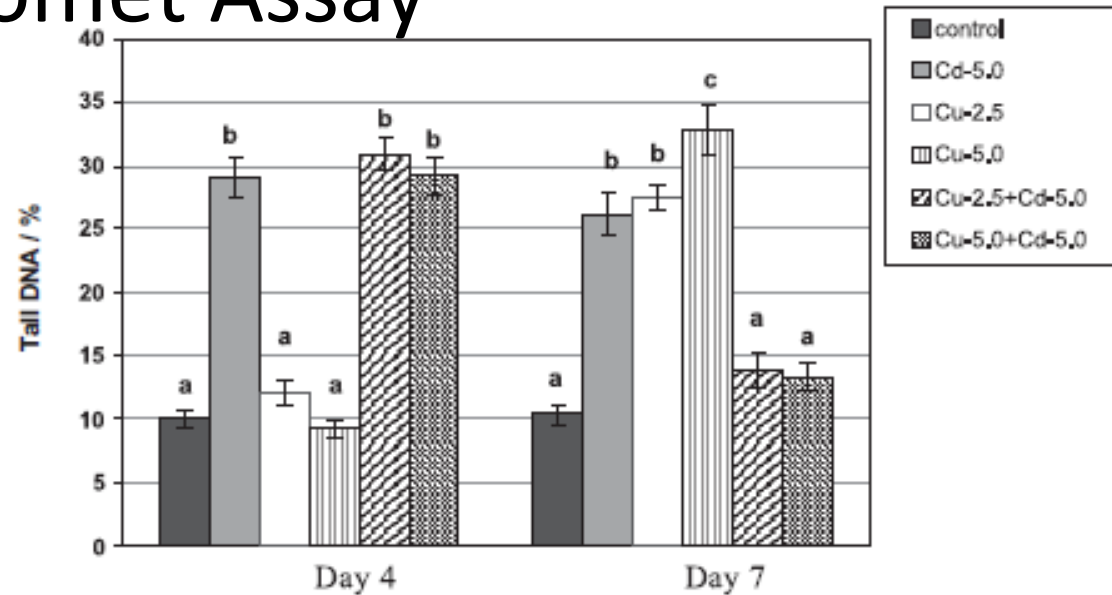
# DNA disruption – Gelelectrophoresis and Comet Assay



1: DNA from control plant, 2:-6: DNA from plants treated with Cd 10, 50, 75, 100, 1000  $\mu$ M

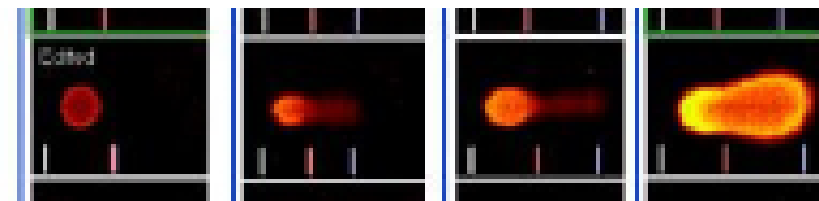
Fojtova & Kovarik, Plant, Cell & Envir. 23:531-537, 2000

## Comet Assay



*Lemna minor* treated with  $\mu$ M concentrations of Cd and Cu. Tail DNA (Comet - length) increase due to metal treatment.

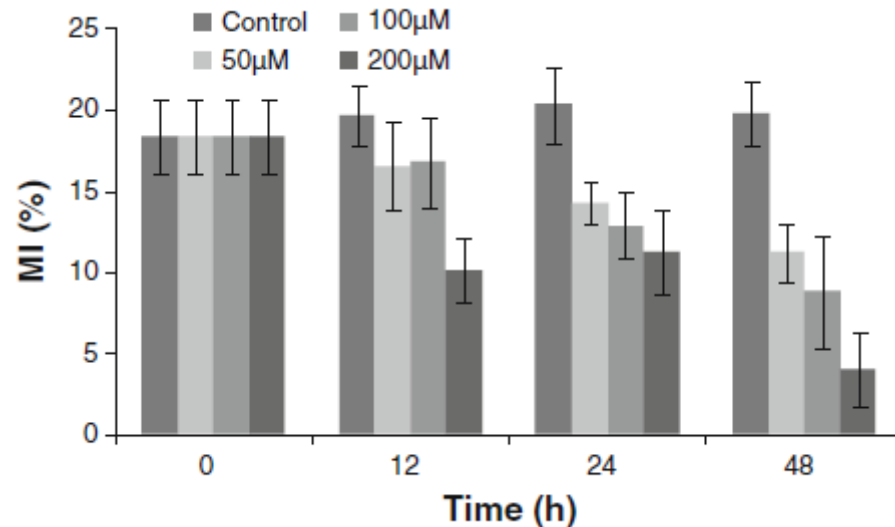
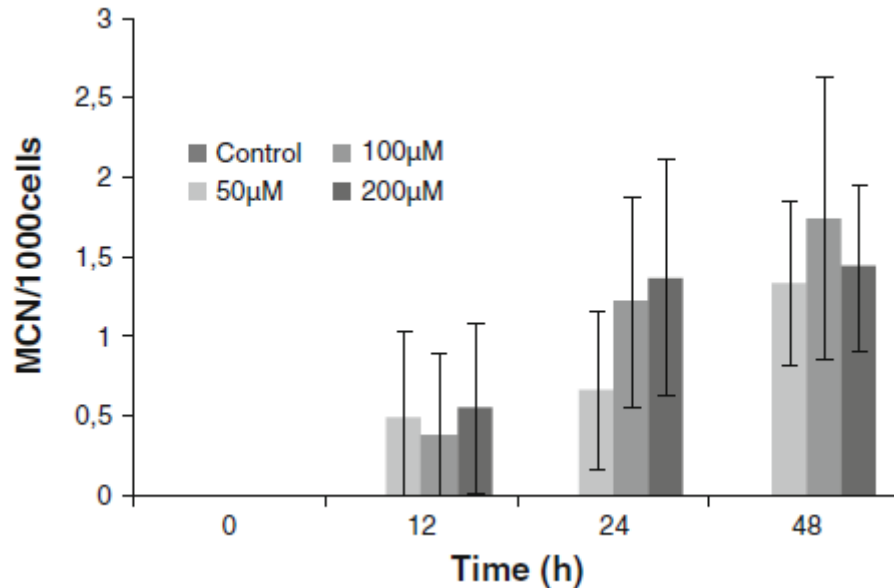
Cvjetko et al., Arh Hig Rada Toksikol 61:287-296, 2010



From the CometAssay Manual, Trevigen<sup>R</sup>

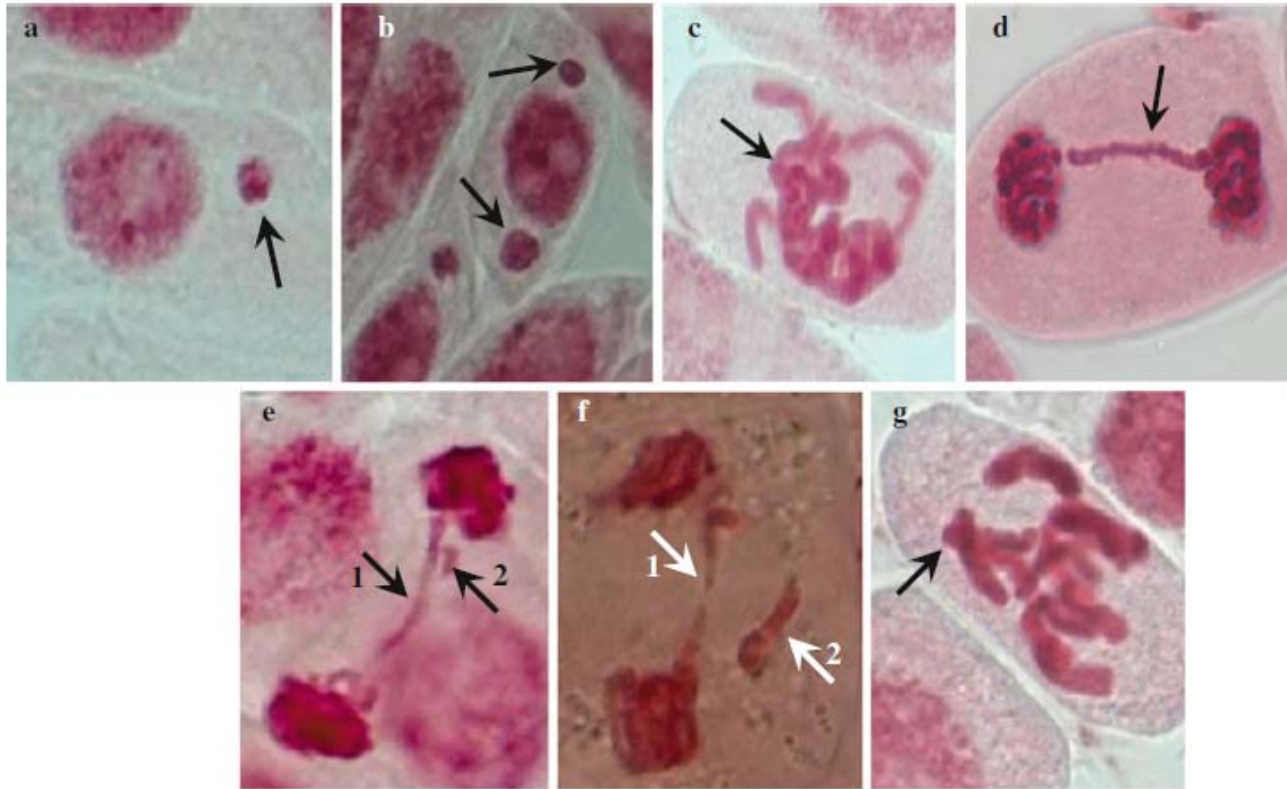


# Micronuclei & Mitotic index



- *Vicia* root meristem cells; established assay
- Micronuclei formation due to malfunctioning cell division
- Dose and time dependent
  - Cd treatment increases MCN
- Mitotic index: ratio of cells in metaphase stage to all cells
  - Cd treatment reduces MI

# Chromosomal aberrations



*Vicia* root meristem cells

a, b, e, f = 50  $\mu$ M Cd

c, d, g = 200  $\mu$ M Cd

a & b = micronuclei

c = sticky chromosome

d = chromosome bridge

e = " " + break

f = " " + isolated

chromosome

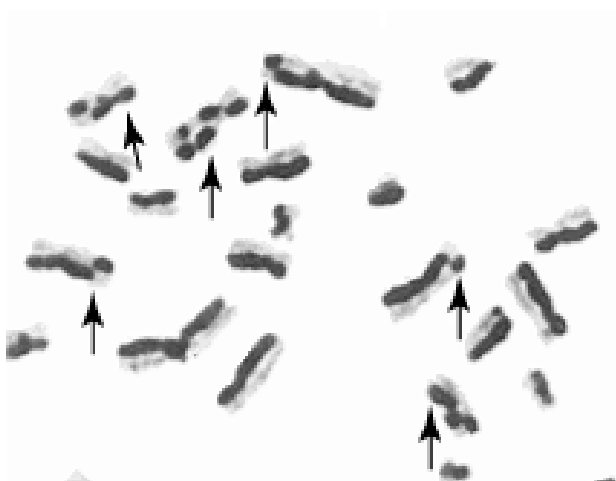
g = lagged chromosome

in metaphase

# Sister chromatid exchange



- Exchange of identical parts of both sister chromatids in the same chromosome after / during DNA replication
- As DNA sequence identical, exchange does not lead to genetic information change ( $\neq$  crossing over)
- Happens in normal cells, but enhanced after treatment with toxic / radioactive substances



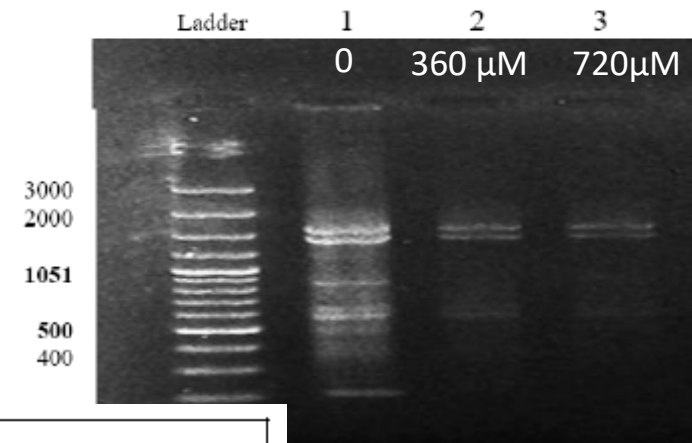
Test substance	Concentration	SCEs/metaphase (mean $\pm$ SE)	MI (mean $\pm$ SE)
Negative Control (Hoagland's nutrient Solution)	0	6.26 $\pm$ 0.29	11.80 $\pm$ 0.11
Cadmium nitrate ( $\mu$ M)	50	6.63 $\pm$ 0.3	9.70 $\pm$ 0.26***
	100	7.43 $\pm$ 0.04**	7.16 $\pm$ 0.32***
	200	7.90 $\pm$ 0.07***	1.16 $\pm$ 0.03***
Positive control (Cyclophosphamide, $\mu$ g/mL)	5	12.11 $\pm$ 0.06***	2.36 $\pm$ 0.31***

\*\* (P < 0.01) and \*\*\* (P < 0.001) compared with negative control.

From: <http://www.siteklabs.com/GenTox/MammalianCellCytogenetics.html>

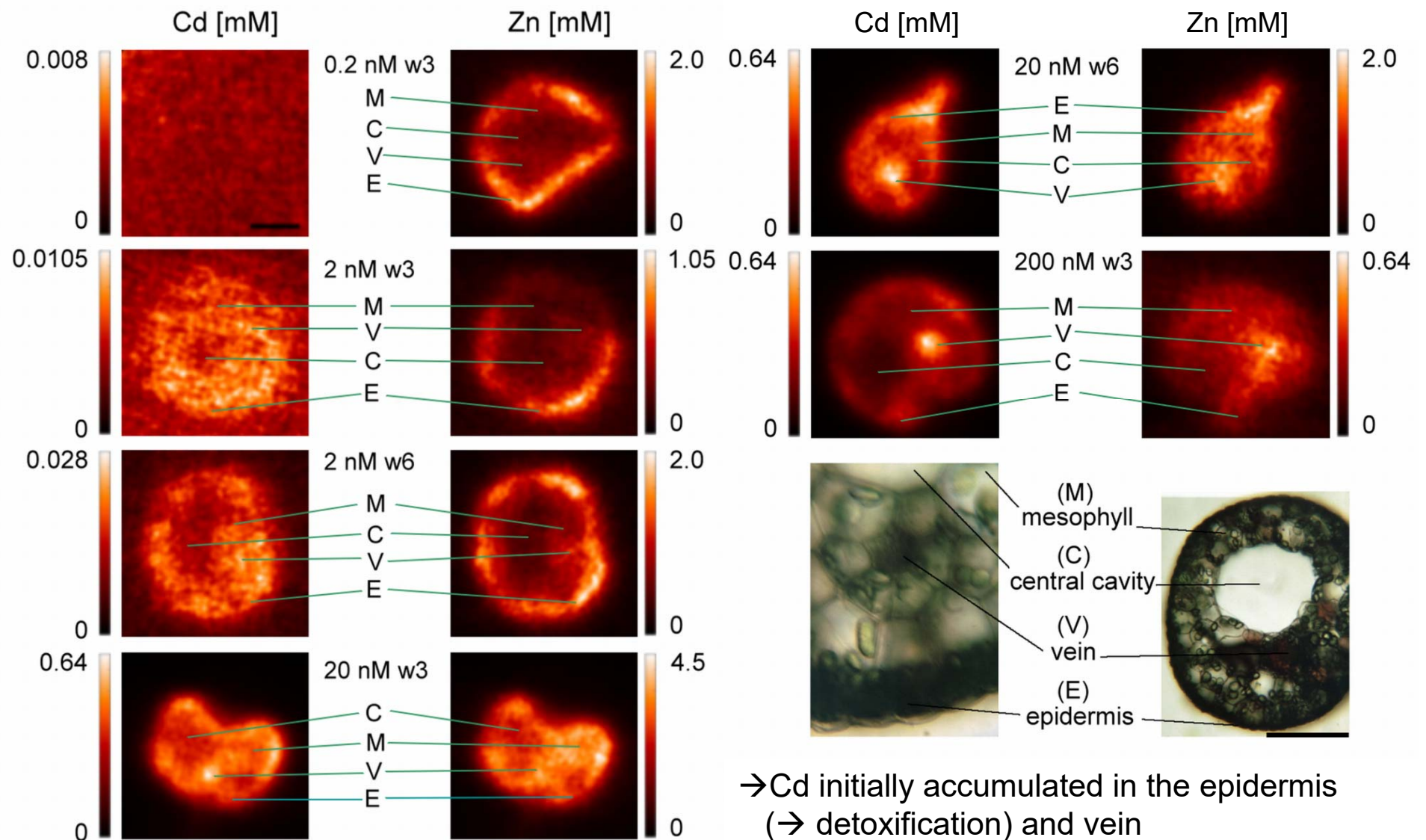
# Random amplification of polymorphism DNA analyses (RAPD)

- Cd interacts with DNA / induces mutation
- New / disappearing primer binding sites
- → New / disappearing bands on gel



Primers names	Treatments				
	Total bands in control	40 (mgL <sup>-1</sup> ) Cd concentration		80 (mgL <sup>-1</sup> ) Cd concentration	
		Appearance of new bands	Disappearance of control bands	Appearance of new bands	Disappearance of control bands
OPA-2	1679, 1500, 1205, 874, 657, 603, 513, 221		1205, 874, 513, 221	900	1205, 874, 513, 221

# Distribution of Cd and its effect on Zn distribution in the non-accumulator shoot model *Ceratophyllum demersum*



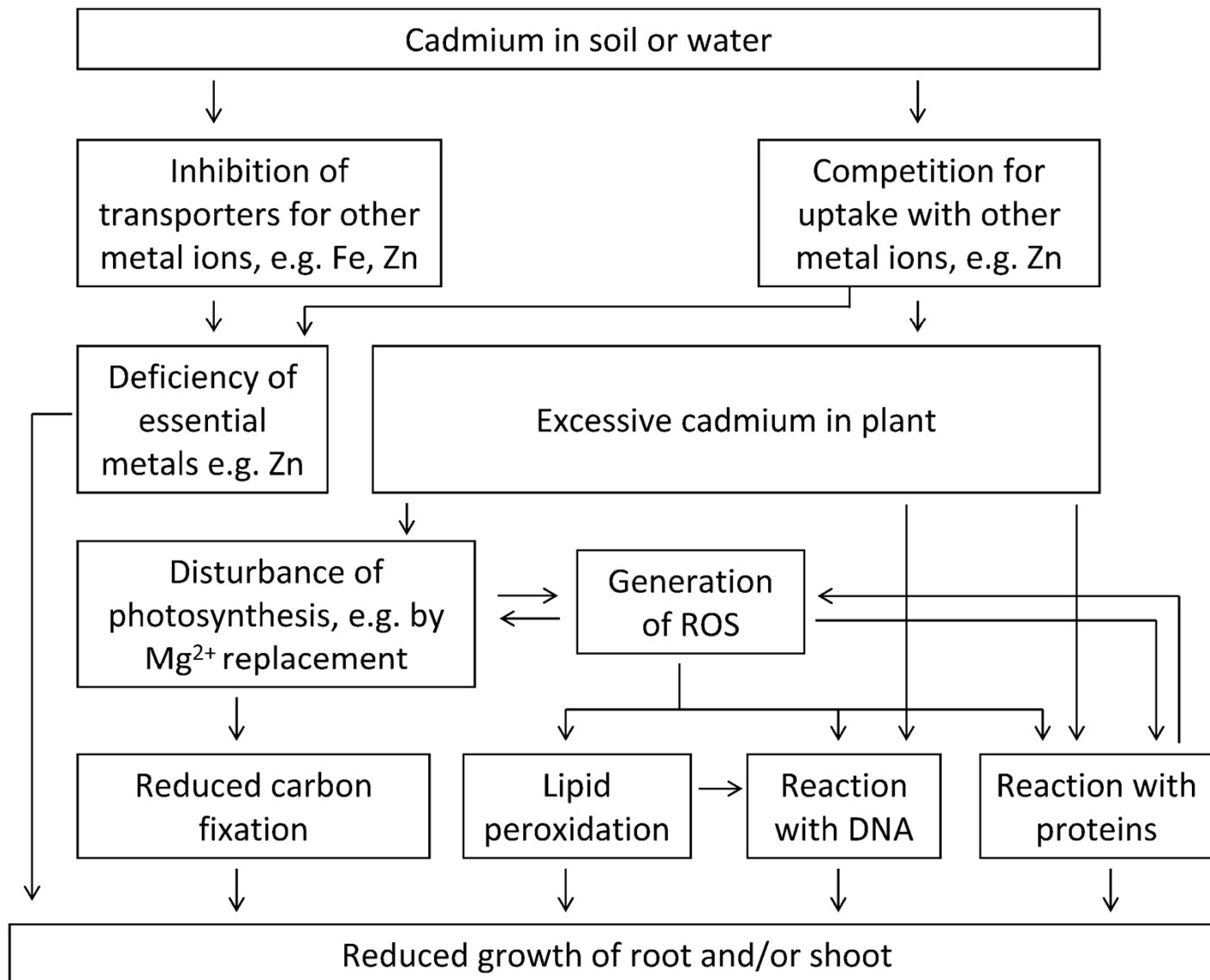
→ Cd initially accumulated in the epidermis  
 (→ detoxification) and vein  
 → lethal Cd inhibited vein export. also of Zn

# Changing distribution of Cd and Zn

Increasing Cd:

- Higher Cd concentration in leaves
- Increased sequestration into non-photosynthetic tissues
- Re-distribution of Zn → inhibited export out of vein





Andresen E, Küpper H (2013) Cadmium Toxicity in Plants. In: Cadmium: From Toxicity to Essentiality, "Metal Ions in Life Sciences Vol. 11;

# Summary Cd toxicity

- Threshold concentration for most toxic effects: 20nM
  - Most fluorescence parameters, growth, pigments
- First site of inhibition: photosynthetic apparatus
  - ROS follow reaction
- Direct inhibition of PSII RC only pronounced in HL
- In LL binding of Cd to LHClI already below 20nM
- Toxicity more pronounced under HL conditions
  - Chl in LHClI acts as buffer under LL

**All slides of my lectures can be downloaded  
from my workgroup homepage**

Biology Centre CAS → Institute of Plant Molecular Biology → Departments  
→ Department of Plant Biophysics and Biochemistry,  
*or directly*

**[http://webserver.umbr.cas.cz/~kupper/AG\\_Kuepper\\_Homepage.html](http://webserver.umbr.cas.cz/~kupper/AG_Kuepper_Homepage.html)**